Protein isolate from the herb, Phyllanthus niruri, protects liver from acetaminophen induced toxicity

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Abstract

Phyllanthus niruri L. (Euphorbiaceae) is a well-known hepatoprotective herbal plant. The aim of this study was to investigate the hepatoprotective action of the protein isolate of Phyl-lanthus niruri (P. niruri) against acetaminophen (APAP) induced hepatotoxicity. Protein iso-late of P. niruri was intraperitoneally injected in mice either prior to (preventive) or after the induction of toxicity (curative). Levels of different liver marker enzymes in serum were measured in normal, control (toxicity induced) and protein isolate treated mice. Protein isolate of P. niruri significantly reduced the elevated Glutamate Pyruvate Transaminase (GPT), and Lactate dehydrogenase (LDH) levels in sera of toxicity induced mice, compared to the control group. Treatment with the protein isolate of P. niruri also enhanced depleted hepatic glutathione (GSH) levels caused by APAP administration. In addition, histological studies supported these results and clearly showed that the protein isolate act as a hepatoprotective agent against APAP-induced toxicity. Combining, data suggest that the protein isolate of P. niruri protects liver tissues against APAP induced hepatic damage.

Introduction

Herbal products have been used since the dawn of civilization on the earth to maintain human health and to get the remedies from various diseases by the vast majority of the world's population. Among these herbs, P. niruri has been widely used all over India for the treatment of jaun-dice and other hepatic disorders [1]. Venkateswaran et al [2] reported that there are one or more materials in P. niruri, which inhibit the replication of woodchuck hepati-tis virus in vivo and decrease the pathological effect of the same virus on woodchuck liver. Moreover clinical studies on humans have also been carried out with P. niruri with-out any side effect [3]. It has been shown in various stud-ies that the herb has potent hepatoprotective activity against various hepatic disorders like viral hepatitis and toxicity caused by different drugs and environmental toxicants [3,4,5]. But mechanisms responsible for this hepatoprotective action of P. niruri, however are not well defined.

Acetaminophen (APAP) overdose is the most frequent cause of drug induced liver injury. The drug is safe at therapeutic doses, but its high dose can cause hepatoxicity in experimental animals and humans [6]. In our laboratory, studies on the hepatoprotective action of P. niruri against APAP toxicity are being carried out. APAP in-toxicated mice were injected with protein isolate of P. niruri, prior to (preventive properties) and after (curative properties) the intoxication. Thereafter, serum levels of marker enzymes related to hepatic integrity, like, GPT, and LDH, as well as cellular GSH level in the liver were determined. In addition, histopathogical studies were done to prove the effectiveness of the protein isolate of P. niruri in the preventive and curative role against APAP induced toxicity in vivo.

Material and methods

Materials

Fresh P. niruri was collected from local market. DTNB (Ellman's reagent), Bradford reagent, and APAP were purchased from Sigma Chemical Co., USA. Kits for the determination of SGPT and ALP were purchased from Span Diagnostics Ltd., India.

Extraction of proteins from the leaves

Fresh leaves were extracted with phosphate buffer, pH-7.4 at 4°C in a cyclomixer. The extract was then centrifuged at 20,000 xg at 4°C. The resultant clear supernatant was then brought to 60% ammonium sulphate saturation and then centrifuged at 20,000 xg at 4°C and the pellet collected was reconstituted in 50 mM phosphate buffer, pH-7.4.

Protein estimation

Different protein concentrations were measured according to the method of Bradford [7].

Animal experiments

Swiss albino mice (male, body weight 20 ± 2 g) were acclimatized under standard laboratory condition for two weeks before starting experiments. They were provided with standard pellet diet and water ad libitum .The animals were divided into several groups of 6 in different models of experiments.

First group served as controls. Second group received only APAP orally at a dose of 300 mg/kg of body weight for 2 days. Third group were pretreated with the protein isolate for 3 days by intraperitoneal injection at a dose of 5 mg/kg body weight of mice and

followed by APAP at the same dose as above. The concentration of protein iso-late to be used was determined by carrying out a dose response study with the protein isolate pretreated group using different doses of protein concentrations ranging from 1- 20 mg/kg of body weight.

Fourth group was post treated with the protein isolate for 3 days after APAP treatment. With this group a time course study was done by injecting P. niruri protein iso-late for 1, 2, 3 and 4 days. Optimum result was obtained when protein was administered for 3 days.

Evaluation of hepatotoxicity

Serum was prepared from the bloods collected. About 200 mg liver tissue was homogenized in 10 volumes of appropriate buffer. The blood serum and supernatant were used to estimate the hepatotoxicity produced by APAP. Serum GPT was measured by 2,4 DNPH method of Reitman and Frankel and LDH [8] was measured using an assay where reduction of NAD to NADH by LDH was measured by monitoring the increase in absorbance at 340 nm. Hepatic GSH level was determined by the method of Ellman [9]. For liver histopathology mice were divided into four groups and treated with APAP and protein isolate as de-scribed earlier. Liver specimens from all the experimental groups were fixed in 10% buffered formalin and sections of about 5 μ m thickness were stained with haematoxylin and eosin stain to study the effect of P. niruri protein iso-late on the APAP treated mice liver.

Statistical analysis

The data were expressed as mean +SEM (n=6), and n refers to the number of animals. Differences between the groups were analysed by one-way ANOVA, using the Epistat package.

Results

Effect of P. niruri protein isolate against APAP induced hepatotoxicity

Dose response study

A dose response study was done by injecting different concentrations of protein isolate prior to APAP administration. The doses used were, ranging from 1-20 mg/kg of body weight of protein. Serum GPT level of all the mice were measured. It is evident that optimum result was obtained at a dose of 5 mg/kg body weight of protein, which was used in all other experiments.

Time dependent effect of P. niruri treatment

A time dependent study was done by injecting protein isolate for 1, 2, 3 and 4 days after 2 days of APAP administration at a dose of 300 mg/kg body weight. Serum GPT levels of these mice were measured. From the results it is evident that 3 days protein injection gave

optimum results. So, further experiments were carried out by injecting the protein fraction for 3 days.

Liver assays

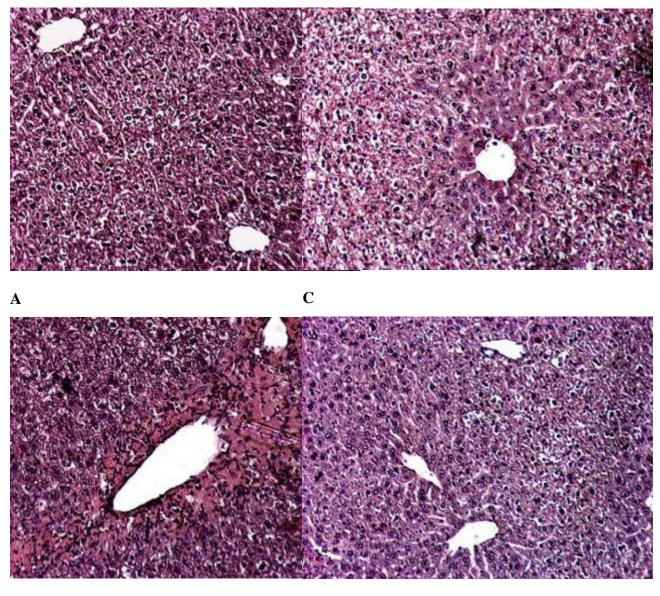
APAP exposure increased serum GPT and LDH levels in APAP treated mice indicating severe liver damage. Protein isolate pretreatment as well as post-treatment for 3 days at a dose of 5 mg/kg body weight significantly reduced the levels of GPT and LDH close to normal level in both the cases. GSH level was depleted to ~30% in APAP treated mice compared to normal mice group. GSH level was restored to almost 75% in protein isolate pretreated mice. When post-treated with protein GSH contents in liver homogenates of APAP treated mice were decreased to ~50% compared to the normal mice group. GSH level of protein isolate post-treated group was increased to ~90%. The data is presented in Table 1.

Table 1: Effect of P. niruri protein isolate on serum GPT, LDH and liver GSH levels during APAP hepatotoxicity. Control- normal mice; APAP- APAP treated mice at a dose of 300 mg/kg body weight; P. niruri + APAP- protein pre-treated for 3 days at a dose of 5 mg/kg body weight and then APAP treated; APAP + P. niruri- APAP treated + pro-tein post treated at a dose of 5 mg/kg body weight for 3 days. Each value represents mean + SEM (n=6). Protein (5mg/ml dose) treated groups were significantly different ($P \le 10-6$) from APAP treated group.

Markers	Control	APAP	P. niruri + APAP	APAP	APAP + P. niruri
SGPT (Units/ml of serum)	25 + 1.2	130 + 6.2*	35. + 3**	62 + 3.1*	28 ± 1.8**
LDH IU/ltr of serum	126.3 + 7.8	282.6 +12.3*	167.7 ± 5.4**	$254.8 \pm 9.9^*$	139 + 6.5**
GSH (nmoles/ mg total protein)	35 + 1.3	12+0.9*	27 + 1.9**	17+0.9*	31.5 + 1.3**

Each value represents mean + SEM (n=6).

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B

D

Fig. 1: Effect of P. niruri protein isolate on liver histopathology.

A. Normal liver

B. Liver from mice treated with APAP at a dose of 300 mg/kg body weight

C. Liver from mice pretreated with protein isolate for 3 days and then PAP treated at a dose of 300 mg/kg body weight for 2 days.

D. Liver from mice treated with APAP at a dose of 300 mg/kg body weight for 2 days and then post-treated with protein isolate.

Histopathological samples were taken from normal, APAP treated and protein isolate treated (both pre and post) mice liver. Histopathological assessment represented in Fig. 1, shows prominent changes including centrilobular necrosis, bile duct proliferation, disorganization of normal radiating pattern of cell plates around central vein etc. in

tissues treated with APAP. Qualitatively, the inflammation in protein isolate treated mice liver before and after APAP administration were less severe compared to that with APAP. These observations significantly prove the protective role of the protein isolate from P. niruri.

Discussion

The liver is a major target organ for toxicity of xenobiotics and drugs as most of the orally ingested drugs pass through the liver and some chemicals are metabolized into toxic intermediate in the liver.

APAP is the most commonly used analgesic and antipyretic and is responsible for more hospitalization after overdose than any other common medication. APAP, when used in high doses, could cause potentially fatal hepatic necrosis [10]. There is general consensus that the formation of an electrophilic metabolite by the P450 system, presumably N-acetyl-p-benzoquinoneimine (NAPQI), is a prerequisite for the injury [11]. NAPQI is effectively detoxified by glutathione to form an acetaminophen-GSH conjugate. After a toxic dose of acetaminophen, total hepatic GSH is depleted [12]. However, after the cellular glutathione stores are depleted, the reactive metabolite covalently binds to a substantial number of cytosolic and mitochondrial proteins [13]. As a consequence, impaired mitochondrial respiration [14], depletion of hepatocellular ATP levels [15], and release of cytochrome c from the mitochondria [16,17] are observed.

The present study demonstrates that the protein isolate of the herb P. niruri has vital hepatoprotective properties against hepatic damage induced by the drug APAP. In our study we treated the mice with APAP at a dose of 300 mg/kg bodyweight. We observed that this resulted in severe hepatic toxicity as evident from the increased levels of liver marker enzymes GPT and LDH. Moreover, APAP treatment reduced hepatic levels of GSH, which is required for the detoxification of NAPQI and induced centrilobular necrosis in liver as evident from histopathological studies. In order to elucidate the protective role of P. niruri against APAP induced hepatotoxicity in mice. It was observed that the protein isolate was able to reduce the increased levels of liver marker enzymes GPT and LDH, significantly. Moreover, it increased the depleted levels of cellular GSH. In histopathological studies, damage around the lobular region of the liver from protein isolate treated mice liver before and after APAP administration was less severe compared to that with APAP alone.

In conclusion, the proteins present in P. niruri could be used as a protective agent against APAP toxicity. Further works are needed to fully characterize the protein(s) present in the herb and elucidate its possible mode of action and that is in progress.

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References

- Syamasundar KV, Singh B, Thakur RS, Husain A, Kiso Y, Hikino H. Antihepatotoxic principles of Phyllanthus niruri herbs. J Ethnopharmacol 1985; 14: 41-44.
- 2. Venkateswaran PS, Millman I, Blumberg BS. Effects of an extract from Phyllanthus niruri on hepatitis B and woodchuck hepatitis viruses: in vitro and in vivo studies. Proc Natl Acad Sci USA 1987; 84: 274-278.
- 3. Thyagarajan SP, Subramanian S, Thirunalasundari T, Venkateswaran PS, Blumberg BS. Effect of Phyllanthus amarus on chronic carriers of hepatitis B virus. Lancet 1988; 2 (8614): 764-766.
- 4. Padma P, Setty OH. Protective effect of Phyllanthus against CCl4-induced mitochondrial dysfunction. Life Sci 1999; 64: 2411-2417.
- 5. Sebastian T, Setty OH. Protective effect of Phyllanthus against ethanol-induced mitochondrial dysfunction. Al-cohol 1999; 17: 29-34.
- 6. Nelson SD. Molecular mechanisms of the hepatotoxicity caused by acetaminophen. Semin Liver Dis 1990; 10: 267-278.
- 7. Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Anal Biochem 1976; 72: 248-254.
- 8. Searey RL. Diagnostic Biochemistry. 1969; McGrane-Hill, New York, NT.
- 9. Ellman GL. Tissue sulphydryl group. Arch Biophy 1959; 82: 70-77.
- 10. Proudfoot AT, Wright N. Acute paracetamol poisoning. Br Med J 1970; 5, 3: 557-558.
- Nelson SD, Bruschi, SA. Mechanisms of acetaminophen-induced liver disease. In Drug-induced Liver Disease. (N. Kaplowitz, L.D. DeLeve, Eds.), 2003; pp. 287– 325. Marcel Dekker, Inc., New York.
- 12. Mitchell JR, Jollow DJ, Potter WZ, Gillette JR, and Brodie BB. Acetaminopheninduced hepatic necrosis: IV. Protective role of glutathione. J Pharmac Exp Ther 1973; 187: 211-217.
- 13. Cohen SD, Pumford NR, Khairallah EA, Boekelheide K, Pohl LR, Amouzadeh HR, and Hinson JA. Selective protein covalent binding and target organ toxicity. Toxicol. Appl Pharmacol 1997; 143: 1-12.
- Meyers LL., Beierschmitt WP, Khairallah EA, Cohen SD. Acetaminopheninduced inhibition of mitochondrial respiration in mice. Toxicol Appl Pharmacol 1988; 93: 378-387.
- 15. Tirmenstein Man, Nelson SD. Acetaminophen-induced oxidation of protein thiols: Contribution of impaired thiol-metabolizing enzymes and the breakdown of adenine nucleotides. J Biol Chem 1990; 265: 3059-3065.
- Adams ML, Pierce RH, Vail ME, White CC, Tonge RP, Kavanagh TJ, Fausto N, Nelson SD, Bruschi SA. Enhanced acetaminophen hepatotoxicity in transgenic mice overexpressing BCL-2. Mol Pharmacol 2001; 60: 907-915.

17. Knight TR, Ho YS, Farhood A, and Jaeschke H. Peroxynitrite is a critical mediator of acetaminophen hepatotoxicity in murine livers: Protection by glutathione. J Pharmacol Exp Therap 2002; 03: 468-475.

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