



RESEARCH ARTICLE



Received on: 16-11-2013 Accepted on: 14-12-2013 Published on: 21-12-2013

Seham M. Botros*

Pharmaceutical Analytical Chemistry Department, Faculty of Pharmacy, Assiut University, Assiut, Egypt Email: seham.botros@gmail.com



QR Code for Mobile users

Conflict of Interest: None Declared !

A novel spectrofluorimetric determination of four anti-TB drugs in their pure and pharmaceutical dosage forms by quenching effect on the fluorescence of NBS-phenothiazine product

Abdel Maaboud I*. Mohamed, Fardous A. Mohamed, Noha N. Atia and Seham M. Botros

Pharmaceutical Analytical Chemistry Department, Faculty of Pharmacy, Assiut University, Assiut, Egypt

Abstract

Simple and sensitive method has been developed and validated for determination of ethambutol (EMB), isoniazid (INH), pyrazinamide (PZA) and rifampicin (RIF) in pure and pharmaceutical dosage forms. The method is based on measuring the quenching effect of studied drugs on the fluorescence intensity of NBS-phenothiazine oxidation product (NBS-Phz) in a buffered medium (pH 7, λ_{ex} 271 and λ_{em} 375 nm). Different factors affecting the reaction were studied and optimized. Under the optimized conditions, linear relationships with good correlation coefficients (0.9995-0.9999) were obtained. The limits of detection were 0.139, 6.39 x 10⁻³, 0.029 and 0.180 µg ml⁻¹ for EMB, INH, PZA and RIF respectively. The precision of the method was satisfactory with relative standard deviation less than 2 %. Good accuracy was assessed with recovery percentages ranged from 97.24 to 101.2 %. The results were favorably compared with those of the official method. Therefore, the developed method provides applicable tool for determination of studied drugs in pure, pharmaceutical formulation and in quality control laboratories.

Keywords: Ethambutol, Isoniazid, Pyrazinamide, Rifampicin, Fluorescence quenching, NBS, Phenothiazine.

Cite this article as:

Abdel Maaboud I. Mohamed, Fardous A. Mohamed, Noha N. Atia and Seham M. Botros. A novel spectrofluorimetric determination of four anti-TB drugs in their pure and pharmaceutical dosage forms by quenching effect on the fluorescence of NBS-phenothiazine product. Asian Journal of Biomedical and Pharmaceutical Sciences; 03 (26); 2013; 21-27.

1. INTRODUCTION

In the last decade, tuberculosis (TB) has remerged as one of the leading causes of death (nearly 3 million deaths annually). A major public health problem worldwide, TB is now global emergency. EMB, INH, PZA and RIF are the first line anti-TB drugs (Fig. 1). The standard treatment for TB is to treat the patient with a combination of these four compounds for two months, followed by INH and RIF alone for an additional four months. For more than 50 years, TB has been treated with combination drug therapy and there are a number of available combination drug products with different drug contents and composition. These compounds are used in combination because they have different modes of action [1-3].

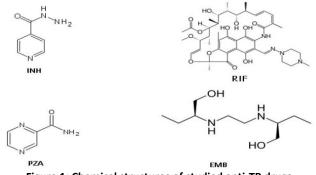


Figure 1: Chemical structures of studied anti-TB drugs Owing to the therapeutic importance of these drugs, numerous different analytical methods have been developed for their quantitative determination in pure, pharmaceutical dosage forms and/or biological fluids. These methods are; liquid chromatography [4-71. thin layer chromatography [8, 9], gas chromatography [10], capillary electrophoresis [11-13], electrochemically [14-16], spectrophotometry spectrofluorimetry [21, [17-20], 221 and chemiluminescence methods [23-25]. These methods, suffer from their sophisticated instrumentation and high-analysis cost. Moreover, these instruments are not available in most quality control laboratories specially, third world countries.

In general, spectrofluorometry is considered one of the most convenient analytical techniques, because of its inherent simplicity, low cost, and wide availability in most quality control laboratories. To the best of our knowledge, there were no reported spectrofluorimetric methods on PZA or one method for the four studied drugs. For these reasons, the present study describes simple, sensitive and economical spectrofluorometric method for the analysis of four anti-TB drugs in their pure and pharmaceutical dosage forms. The proposed method is based on measuring the quenching effect of these drugs in buffered medium (pH 7) on the fluorescent

reaction product between NBS and phenothiazine. **2. EXPERIMENTAL**

2.1. Apparatus:

RF-5301 PC spectrofluorimeter (Shimadzu, Japan), with 1-cm matched quartz cells, were used for all measurements. The spectrofluorimeter was set at excitation and emission slit width of 3 nm.

2.2. Materials and reagents:

RIF and INH working reference standards were gifted from Pharco, Cairo, Egypt. PZA working reference standard was kindly supplied from Amoun, Cairo, Egypt and EMB by Memphis/ Zoga, Cairo, Egypt. Phenothiazine (Sigma chemical Co., USA) freshly prepared ethanolic solution (5×10^{-4} M). Nbromosuccinimide (Merck, Germany) freshly prepared aqueous solution (10^{-3} M). Teorell and Stenhagen

Pharmaceutical preparations	Ingradients	Nominal content, mg	Manufacturer
Etibi® 500 tablets	EMB	500 mg/tablet	Memphis/ Zoga, Cairo, Egypt
Isocid fort® tablets	INH	200 mg/tablet	Cid, Cairo, Egypt
Rifampicin® capsule	RIF	300 mg	El Nasr Pharm. Chem. Co., ADWIC
Rimactane® capsule Rimactane® suspension	RIF	300 mg 20 mg/ml	Novartis Pharm. Co.
Rifactine® capsule Rifactine® suspension	RIF	300 mg 2 g/100 ml	Medical Union Pharm. Co., (MUP)
Rifadin [®] capsule	RIF	300 mg	Nile Co. for Pharm. & Chem. Ind.
P.T.B tablets®	PZA	500 mg	Amoun Pharm. Co.

Table 1: Commercial pharmaceutical preparations analyzed bythe proposed spectrofluorimetric method

buffer solution [26] of the pH range 2-10 was prepared in freshly boiled and cooled distilled water. The buffer composed of phosphoric acid, citric acid, 1 M sodium hydroxide, adjusted to the required pH with 0.1 M hydrochloric acid.

All solvents and other chemicals used throughout this study were of analytical grade. Double distilled water has been used.

2.2.1. Pharmaceutical dosage forms:

Different pharmaceutical formulations analyzed (Table 1), were purchased from the local market.

2.3. Preparation of solutions:

2.3.1. Stock standard solutions:

An accurately weighed amount (25 mg) of each studied drug was transferred into a 25-ml volumetric flask. The

powder was dissolved in 10-ml methanol. The solution was then diluted to the mark with the same solvent to obtain a working standard solution of 1 mg ml⁻¹ of each of the studied drugs. Further dilutions were made to obtain the suitable drug concentrations; 1-4, 0.1-0.35, 0.1-1 and 1-5 μ g ml⁻¹. These solutions were found to be stable for at least 1 weak when kept in the refrigerator. **2.3.2. Sample preparation:**

Tablets

Ten tablets of each formulation were weighed and finely powdered. A quantity of the mixed powder equivalent to 25 mg of each drug was transferred into a 25-ml calibrated flask, dissolved in 10-ml methanol, swirled and sonicated for 10 min, completed to volume with the same solvent, shaken well for 10 min, and filtered. The procedure was then completed as described for preparation of stock standard solutions.

Capsules

The contents of ten capsules were evacuated and accurately weighed and finely powdered. A portion of the powder equivalent to 25 mg of RIF was weighed and quantitatively transferred into a 25 ml volumetric. The powder was then dissolved in a suitable volume of methanol, shacked and sonicated for 10 min. The solution was then filtered to be used in further experiments.

Suspension

A suitable volume was quantitatively transferred (equivalent to 25 mg of RIF) into a 25 ml volumetric flask containing 10 ml methanol. The solution was shacked and sonicated for 10 min., and then the volume was made up to the mark with the same solvent. The solution was then filtered to give a concentration of 1 mg ml⁻¹.

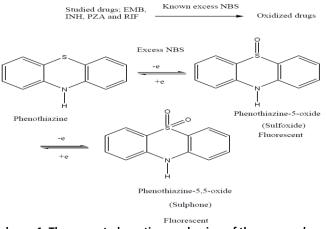
2.4. Determination procedure:

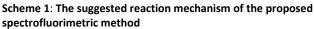
Accurately transfer one milliliter of standard or sample solution into 10 ml volumetric flask; add one milliliter of the buffer pH 7 and mix well. Then add one milliliter of 10⁻³ M NBS. Allow the mixture to stand for 15 min. and finally add 1 ml of 5 x 10⁻⁴ M phenothiazine stand at ambient temperature for 5 min. complete the solution to the mark with distilled water. Measure Δ RIF at λ_{em} 375 nm (λ_{ex} 271 nm) against blank treated similarly.

3. RESULTS AND DISCUSSIONS

3.1. Method development:

N-bromosuccinimide has been extensively used as brominating and oxidizing agent for many organic compounds [27, 28]. In the present work, each studied drug was treated with excess amount of NBS. The excess of NBS was then determined using phenothiazine reagent which undergoes oxidation to the fluorescent products (sulfoxide or sulphone) [29]. These products could be measured fluorimetrically at λ_{ex} 271 nm and λ_{em} 375 nm according to scheme 1. Fig. 2 illustrates the excitation and emission spectra of the studied drugs (3 μg ml $^{\rm 1}$) and their quenching effects.





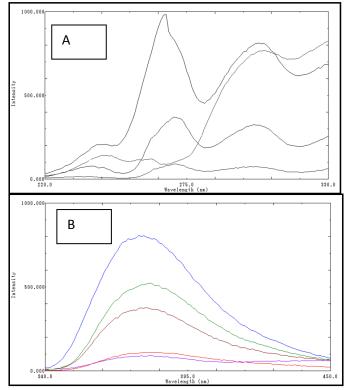


Figure 2: a) Excitation and b) Emission spectra of NBS-Phenothiazine product in absence (blank) and presence of studied drugs

3.2. Optimization of reaction conditions:

All factors affecting the reaction were optimized by altering one variable and keeping the others constant by using 10 $\mu g~ml^{\rm -1}$ of each studied drug.

These factors include:

3.2.1. NBS concentration:

The effect of different concentrations of Nbromosuccinimide (10^{-5} to 10^{-2} M) were tested for the effect on Δ RFI of NBS-Phz product (Fig. 3). It was found that maximum relative fluorescence intensity difference obtained upon using 10^{-3} M NBS.

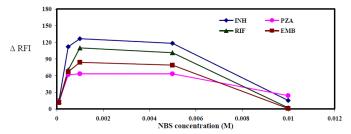


Figure 3: Effect of NBS concentration (M) on Δ RFI of the produced oxidized phenothiazine in presence of drugs under investigation 3.2.2. Phenothiazine concentration:

Various concentrations of phenothiazine ranged from 10⁻⁵ to 10⁻² M were examined. Maximum response was obtained upon using 5 x 10^{-4} M phenothiazine in ethanol (Fig.4).

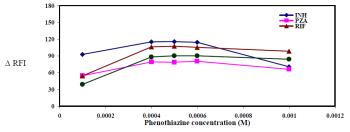


Figure 4: Effect of phenothiazine concentration (M) on Δ RFI of NBS-Phenothiazine product in presence of anti-TB drugs

3.2.3. pH:

For investigating the effect of pH, the reaction was performed at different pH values (2-10). The results indicated that the fluorescence intensity difference was pH dependent (Fig.5.). The optimum pH was found to be 7.0.

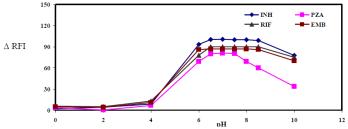


Figure 5: Effect of pH on Δ RFI of NBS-Phenothiazine product in presence of studied drugs

3.2.4. Reaction time and temperature:

It was found that high temperature leading to unstable oxidized form of phenothiazine according to the practical study (Fig. 6).

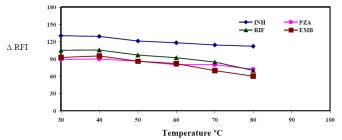


Figure 6: Effect of temperature on Δ RFI measured at λ_{ex} 271 and λ_{em} 375 nm

Therefore reaction at ambient temperature was selected. Fifteen minutes was selected as the reaction time of choice between each drug and NBS.

3.2.5. Reaction time between phenothiazine and NBS:

Reaction time between NBS and phenothiazine was studied for anti-TB drug through 25 min. Five minutes standing was chosen before final dilution with distilled water. The stability time of the oxidized product was studied by carrying out the general assay procedure varying standing time from 5 to 60 min. It was observed that the produced relative fluorescence intensity difference of the reaction product remained stable at least 1 hr.

3.2.6. Diluting solvent:

In order to select the most appropriate solvent for dilution, different solvents were tested: water, methanol. ethanol. dimethylformamide and dimethylsulphoxide. Water was found to be an ideal diluting solvent (Table 2) as it afforded maximum sensitivity, and therefore it was selected for further investigations.

		$\Delta \text{ RFI}^{a}$			
Solvent	$\lambda_{ex}/\lambda_{em}$ (nm)	EMB	INH	PZA	RIF
Water	271/375	101	127	74	107
Methanol	272/370	94	103	72	99
Ethanol	274/371	92	64	30	97
DMSO	275/376	82	64	41	85
DMF	274/369	77	59	41	82

Table 2: Effect of different solvents on $\lambda_{ex}/\,\lambda_{em}$ and Δ RFI caused by the reaction of studied drugs with NBS and phenothiazine

^a Values for all solvents are mean of three determinations; the RSD for the readings were < 2%

3.3. Validation of the proposed method:

The method was validated according to International Conference on Harmonization guidelines (ICH) on the validation of analytical methods, in terms of linearity, precision, limit of detection, limit of quantitation, robustness, accuracy and analysis of marketed formulations [30].

3.3.1. Linearity and sensitivity:

Under the specified optimum reaction conditions, the calibration curves for anti-TB drugs were constructed by measuring a series of six concentrations of the standard solutions. All measurements were carried out using six replicate measurements (n=6). The assays were carried out according to the general procedure previously established for each drug. In all cases, standard curves were linear with acceptable intercepts and very good correlation coefficients in the general concentration range of 1-4, 0.1-0.35, 0.1-1 and 1-5 µg ml⁻¹ for EMB, INH, PZA and RIF respectively (Table 3). The LOD was 0.139, 6.39 x 10⁻³, 0.029 and 0.180 µg ml⁻ ¹, while the LOQ was 0.420, 0.019, 0.088 and 0.546 μ g

ml⁻¹ for EMB, INH, PZA and RIF respectively. Limits of detection (LOD) and limits of quantitation (LOQ) were determined using the formula: LOD or LOQ = kSD_a/b , where, k = 3.3 for LOD and 10 for LOQ, SD_a is the standard deviation of the intercept, and b is the slope.

Parameter	EMB	INH	PZA	RIF
Linearity range (µg ml ⁻ ¹)	1-4	0.1-0.35	0.1-1	1-5
Correlation coefficient (r)	0.9995	0.9999	0.9996	0.9996
Intercept ±	-2.23 ±	-6.9 ±	-0.644 ±	6.9 ±
SD*	0.336	0.275	0.657	0.546
Slope ± SD*	0.008 ±	0.142 ± 1	0.075 ± 1	0.010 ±
	1.3 x 10 ⁻⁵	x 10-3	x 10-3	1.6 x 10 ⁻⁵
LOD (µg ml-1)	0.139	6.39 x 10 ⁻ 3	0.029	0.180
LOQ (µg ml-1)	0.420	0.019	0.088	0.546

Table3:Quantitativeparametersfortheproposedspectrofluorometric method for the analysis of studied anti-TBdrugs

* Mean of six replicates

3.3.2. Precision:

The precision of the method was estimated by measuring six replicate samples of each drug within the same day and on different days. Three concentrations were selected which covers low, medium and high levels to the calibration curves (1, 2, 4), (0.1, 0.2, 0.3), (0.2, 0.4, 0.6) and (2, 3, 4) μ g ml⁻¹ for EMB, INH, PZA and RIF respectively. The assays gave satisfactory results ranged from 98.18 to 101.5 with relative standard deviation less than 2 % (Table 4). This level of precision of the proposed method was adequate for the quality control analysis of the first line anti- TB drugs in their pharmaceutical dosage forms.

Authentic drug	Concentration (µg ml-1)	Intra-day precision % Found ± RSD*	Inter-day precision % Found ± RSD*
EMB	1 2 4	98.51 ± 1.20 99.73 ± 1.75 100.4 ± 0.71	98.91 ± 1.40 99.53 ± 1.20 98.75 ± 0.88
INH	0.1 0.2 0.3	$\frac{101.3 \pm 0.92}{98.18 \pm 0.79}$ 99.90 ± 0.66	98.42 ± 0.97 101.5 ± 1.03 99.66 ± 0.71
PZA	0.2 0.4 0.6	98.67 ± 1.79 98.44 ± 1.29 98.58 ± 0.87	100.3 ± 1.96 98.96 ± 0.93 98.76 ± 0.82
RIF	2 3 4	99.75 ± 1.42 98.33 ± 0.64 99.00 ± 0.90	98.85 ± 0.90 98.88 ± 0.98 100.6 ± 0.77

 Table
 4: Intra-day and inter-day precision of the proposed spectrofluorimetric method at three concentration levels

 * Standard doubtion of file concentration
 Secondard doubtion of file concentration levels

* Standard deviation of six replicates

3.3.3. Robustness:

Robustness of the procedure was assessed by evaluating the influence of small variation in experimental variables: NBS and phenothiazine concentrations, pH, drug-NBS and NBS-phenothiazine reaction times on the analytical performance of the method. In these experiments, one experimental parameter was changed while the other parameters were kept unchanged, and the recovery percentage was calculated each time (Table 5). The small variations in any of the variables didn't significantly affect the results where the percentage recovery ranged from 96.15 to 104.5 with SD less than 2 %. This gave an indication for the reliability of the proposed method during routine work.

	% Recovery ± SD*			
Variable studied	EMB ^a	INH ^b	PZA ^c	RIF ^d
NBS concentration				
0.0011 M	100.6 ±	97.65 ±	97.61 ±	104.5 ±
0.0009 M	1.23	1.13	1.38	0.58
	96.54 ±	96.95 ±	97.95 ±	98.92 ±
	0.47	1.02	1.29	0.57
Phenothiazine				
concentration	100.8 ±	100.5 ±	97.15 ±	100.9 ±
0.00052 M	1.67	0.40	0.58	1.00
0.00048 M	97.88 ±	99.89 ±	97.92 ±	96.33 ±
	1.39	1.76	1.01	1.49
pH				
7.5	99.62 ±	98.12 ±	97.59 ±	99.91 ±
6.5	1.42	0.89	0.77	0.99
	100.8 ±	99.06 ±	98.06 ±	99.98 ±
	0.93	0.89	0.87	1.09
Reaction time between drug				
and NBS	102.2 ±	97.62 ±	96.37 ±	104.4 ±
17 min.	1.22	0.39	0.70	0.52
13 min.	100.6 ±	99.06 ±	96.15 ±	100.6 ±
	0.94	0.73	0.34	1.15
Reaction time between NBS				
and phenothiazine	101.4 ±	99.88 ±	98.26 ±	102.5 ±
6 min.	0.46	1.13	0.69	1.10
4 min.	98.41 ±	96.59 ±	97.48 ±	96.98 ±
	0.46	0.73	1.20	1.69

Table 5: Influence of small variation in the assay conditions on the analytical performance of the proposed method for analysis of the four anti-TB drugs

* Average of three determinations.

 ${}^{a,\,b,\,c,\,d}$: the concentration of drugs used were 2, 0.2, 0.4 and 3 μg ml-1 respectively

3.3.4. Analysis of pharmaceutical dosage forms and accuracy testing:

The available pharmaceutical dosage forms were subjected to the analysis by the proposed, as well as the official methods [31-34] and the obtained results were statistically compared with each other. The mean percentage recoveries relative to the labeled amounts, obtained by the proposed method ranged from 97.24 to 101.2 % (Table 6). With respect to t- and F-tests, no significant differences were found between the calculated and theoretical values of both the proposed and official methods at 95% confidence level. This indicated similar accuracy and precision in the analysis of studied drugs in tablets (Table 6). Also, the accuracy was checked by using standard addition method [35]. A fixed weight of tablet or capsule equivalent to 25 mg of each drug was used, then (5, 10 and 15 mg) of pure drug were added separately.

	Proposed method	Official method % Recovery ± SD ^a	
Dosage form	% Recovery ± SD ^a		
Etibi [®] 500 tablets	99.53 ± 1.82 ^b	96.79 ± 1.06	
	<i>t</i> = 2.26		
	F = 2.94		
Isocid fort [®] tablets	97.24 ± 0.97	98.15 ± 1.54	
	<i>t</i> = 0.86		
	F = 2.55		
P.T.B [®] tablets	98.96 ± 1.25	99.64 ± 2.63	
	<i>t</i> = 0.30		
	F = 4.44		
Rifadin [®] capsule	101.2 ± 2.02	100.5 ± 1.46	
	t = 0.47		
	F = 1.91		
Rifampicin [®] capsule	99.91 ± 0.99	99.31 ± 1.94	
	<i>t</i> = 0.48		
	F = 3.86		
Rimactane [®] capsule	100.2 ± 2.23	98.85 ± 1.36	
	<i>t</i> = 0.91		
	F = 2.70		
Rifactine [®] capsule	97.64 ± 1.12	98.89 ± 2.39	
	<i>t</i> = 0.82		
	F = 4.54		
Rifactine [®] suspenion	100.2 ± 1.51	99.76 ± 2.32	
	<i>t</i> = 0.30		
	F = 2.36		
Rimactane®	98.60 ± 1.49	98.17 ± 1.27	
suspension	<i>t</i> = 0.39		
	F = 1.37		

Table 6: Determination of studied anti-TB drugs in their dosage forms by the proposed spectrofluorimetric and official methods

* Average of three determinations

** Theoretical values for t and F at 95 % confidence limits (t = 2.78) and (F = 19) respectively

		1		
Drug/ dosage	Added	Found(mg)	%	SD*
form	(mg)		Recovery	
	5	4.97	99.40	2.00
EMB/ Etibi	10	9.78	97.80	1.44
tablets	15	15.3	102.0	1.83
	5	5.02	100.4	1.22
INH/ Isocid fort	10	9.98	99.80	1.76
tablets	15	15.2	101.1	1.00
	5	5.02	100.4	1.00
PZA/ P.T.B tablets	10	10.02	100.2	1.85
	15	14.92	99.47	0.56
	5	5.10	102.0	1.39
RIF/ Rimactane	10	10.2	102.0	1.73
capsule	15	14.9	99.33	1.83

Table 7: Recovery studies for the determination of anti-TBdrugs using standard addition method by the proposedspectrofluorimetric method

* Average of five determinations

4. CONCLUSION

The present study described a validated spectrofluorimetric method for the analysis of four antituberculous drugs in their available dosage forms. The method was simple, rapid, accurate, and reliable for the determination of studied drugs without interference from the common excipients, the proposed method is of great value in quality control analysis of these drugs owing to its improved simplicity, sensitivity, low-cost, and its independence on expensive instruments, or critical analytical reagents.

5. REFERENCES

[1] Manfred E, Burger's medicinal chemistry and drug discovery, Fifth ed., vol. II: 576-586.

[2] Jaime N & William A, *Text* book of organic medicinal and pharmaceutical chemistry, Tenth ed., 204-207.

[3] Perry B, Raymond W, The pharmacological basis of therapeutics, Ninth ed., chapter 48, 1155, 1161, 1162.

[4] Ken-ichi M, Fumio I, Mitsuo W, Determination of isoniazid, acetylisoniazid and isonicotinic acid in human urine by HPLC coupled with post column photochemical reaction and fluorescence detection, Anal. Sci., 1990; 6: 515-518.

[5] Benetton S A, Kedor-Hackmann E R M, Santoro M I R M, Borges V M, Reversed phase high performance liquid chromatographic determination of rifampicin in the presence of its acid-induced degradation products, J. Liq. Chrom. & Rel. Technol., 1998; 21(20): 3215-3221.

[6] Khuhawar M Y, Rind F M, High performance liquid chromatographic determination of isoniazid, pyrazinamide and rifampicin in pharmaceutical preparations, Pak. J. Pharm. Sci, 1998; 11(2): 49-54.

[7] Kumud S, James D, Suresh K, Ramesh N, Sasijith, SL, Method development and validation of ethambutol in human urine by using LC-MS/MS, Inter. J. Res. Pharm. & Biosci., 2011 ; 1(1):1-6.

[8] Guermouche S, Guermouche M H, Solid-phase extraction and HPTLC determination of isoniazid and acetyl isoniazid in serum, comparsion with HPLC, J. Chrom. Sci., 2004; 42:250-255.

[9] Ali J, Ali N, Sultana Y, Baboota S, Faiyaz, S, Development and validation of stability-indicating HPTLC method for analysis of antitubercular drugs, Acta Chromatographica, 2007; 18:168-179.

[10] Khuhawar M Y, Zardari L, Ethyl chloroformate as a derivatizing reagent for the gas chromatographic determination of isoniazid and hydrazine in pharmaceutical preparations, Anal. Sci., 2008; 24(11):1493-1496.

[11] Tsai I, Liu H, Kuo P, Wang J, Shen L, Kuo C, Quantitative determination of isoniazid in biological samples by cation-selective exhaustive injection sweeping-micellar electrokinetic chromatography, Anal. Bioanal. Chem., 2011; 401:2205-2214.

[12] Liu Y, Fu Z, Wang L, Capillary electrophoresis analysis of isoniazid using luminol-periodate potassium chemiluminescence system, Luminescence J., 2011; 26:397-402.

[13] Faria A F, Vasconcelos J P, Goncalves R B, De souze N V M, De Oliveira A L M, Simultaneous analysis of isoniazid and its impurities by capillary zone electrophoresis, Chromatographia, 2012; 75:1335-1339.

[14] Hahn Y, Shin S, Electrochemical behavior and differential pulse polarographic determination of rifampicin in the pharmaceutical preparations, Arch. Pharm. Res. J., 2001; 24(2):100-104.

[15] Atta N F, Galal A, Ahmed R A, Voltammetric behavior and determination of isoniazid using PEDOT electrode in presence of surface active agents, Int. J. Electrochem. Sci., 2011; 6:5097-5113.

[16] Bergamini M F, Santos D P, Zanoni M V B, Electrochemical behavior and voltammetric determination of pyrazinamide using a poly-histidine modified electrode, J. Electroanal. Chem., 2013; 690(1):47-52.

[17] Mohamed E M, Sensitive spectrophotometric method for determination of non UV absorbing ethambutol, Anal. Lett., 1992, 25(2):269-280.

[18] Chenna G P, Shetty S K, Pai J B, Gopinath B, Ahmed M, Development of spectrophotometric methods for the estimation of pyrazinamide in bulk and pharmaceutical formulations, Inter. J. ChemTech Res., 2011; 3(2):737-741.

[19] Al-Enizzi M S, Al-Sabha T N, Al-Ghabsha T S, Use of charge

transfer complex formation reaction in spectrophotometric microdetermination of some drugs, Jordan J. Chem., 2012; 7(1):87-102.

[20] Khamar J C, Patel S A, First derivative spectrophotometric method for the simultaneous estimation of rifampicin and piperine in their combined capsule dosage form, Asian J. Pharm. & Life Sci., 2012; 2(1):49-55.

[21] Garcia Bautista J A, Garcia Mateo J V, Martinez Calatayud J, Spectrofluorimetric determination of iproniazid and isoniazid in a flow injection system provided with a solid phase reactor, Anal. Lett., 1998; 31(7):1209-1218.

[22] Kamat B P, Seetharamappa J, Mechanism of interaction of vincristine sulphate and rifampicin with bovine serum albumin: Aspectroscopic study, J. Chem. Sci., 2005; 117(6):649-655.

[23] Zheng X, Guo Z, Zhang Z, Flow injection electrogenerated chemiluminescence determination of isoniazid using luminal, Anal. Sci., 2001; 17:1095-1099.

[24] Yong MA, Zhang B, Zhao L, Guo G, Lin J, Determination of rifampicin by peroxomonosulphate-cobalt (II) chemiluminescence system, Chin. J. Chem., 2008; 26(5):905-910.

[25] Prior JA, Santos JL, Lima JL, Automated chemiluminometric screening of counterfeit drugs of the antituberculosis agent pyrazinamide, J. AOAC Int., 2009; 92(3):830-836.

[26] Pesez M, Bartos J, Colorimetric and Fluorimetric Analysis of Organic Compounds and Drugs, Marcel Dekker Inc., New York, 1974; 628.

[27] Halvatzis S A, Timotheou-Potamia M M, Calokerinos A C, Analyst, 1990; 115, 1229.

[28] Michalowski J, Kojlo A, Talanta, 2001; 854, 107.

[29] Fardous A M, Anal. Lett., 1995; 28, 2491-2501.

 $\left[30\right]$ International Conference on Harmonization (ICH) Topic Q_2

(R₁): Validation of Analytical Procedures: Text and Methodology, Nov. 2005; http/www.ich.org/LOB/media/MEDIA 417. Pdf.

[31] The British pharmacopoeia, Fifth ed., HM stationary office, London, I, 2007; 806, 1128.

[32] The British pharmacopoeia, Fifth ed., HM stationary office, London, II, 2007; 1771, 1809.

[33] The United States pharmacopoeia, the National Formulary, 26th ed., Us pharmacopoeia convention, Washington, D.C, II, 2008; 2115, 2116, 2459, 2460.

[34] The British pharmacopoeia, Fifth ed., HM stationary office, London, III, 2007; 2683, 2879.

[35] Harvey D, Modern Analytical Chemistry, McGraw-Hill, Boston, MA, 2000; 108-114.