



e-ISSN: 2249-622X

RESEARCH ARTICLE

Phytoconstituents isolated from *Diospyros oocarpa* Thwatist

M J Amar dev¹, N Rajarajeshwari²

1Research Scholar, Shri Jagdish Prasad Jhabarmal Tibrewala University, Rajasthan

2Visveswarapura Institute of Pharmaceutical Sciences, Bangalore

Received:
29th April 2013
Received in revised form:
3rd May 2013
Accepted:
6th May 2013
Available online:
12^h May 2013



Online ISSN 2249-622X
<http://www.jbiopharm.com>

ABSTRACT

The chemical examination of the roots of *D. oocarpa* afforded ten compounds on column chromatography and repeated crystallizations, Lupeol, 5-Hydroxy-4-methoxy-2-naphthaldehyde, 4-Hydroxy-5-methoxy-2-naphthaldehyde, 4-Hydroxy-3, 5-dimethoxy-2-naphthaldehyde, β -sitosterol, Plumbagin, Betulinaldehyde, Diospyrin, 8'hydroxyisodiospyrin, Umbelliferone. Isolation of umbelliferone belongs to coumarin is the first time record from this species and also from *Diospyros* genus. Occurrence of naphthoquinones is very common in *Diospyros* species. The author could isolate these quinines from this species also and thus justified the species. Distribution of naphthaldehydes in nature is very rare and this report of occurrence is new to the literature

Keywords: *Diospyros oocarpa*, Habibone, Diospyrin, umbelliferone, 8'hydroxyisodiospyrin, Betulinaldehyde

1. INTRODUCTION

The genus *Diospyros*, belongs to the family Ebenaceae, approximately consists of 450 species and is wide spread, chiefly in tropics and sub tropics¹⁻⁴. About 90 species occur in India comprising mostly trees and rarely shrub. *Diospyros* species are known to elaborate a series of naphthoquinones and pentacyclic triterpenoid saponins. Phytochemical investigation of more than 130 *Diospyros* species led to the isolation of variety of compounds, the majority of which are triterpenoids naphthoquinones and flavonoids. Then literature revealed that these plants also contain pentacyclic triterpenes and juglone based naphthoquinones⁵⁻⁸. This genus is so interesting that almost all plants were found to possess therapeutic properties⁹. Totally, *Diospyros* genus is economically and medicinally the most important genus of Ebenaceae. About 41 species are indigenous to India, grown mostly in Western Ghats of Karnataka, Goa and Maharashtra, forests of Deccan, Assam and Bengal, and a few are in North India. Medicinally, *Diospyros* species are used as anthelmintic, anti-inflammatory, antibacterial, antifungal, antioxidant, anticancer, antiviral, molluscicidal, piscicidal and termite resistant activities¹⁰⁻¹³.

D. oocarpa Thwatist (Syn: *D. marmorata*) is a moderate sized tree with shining leaves. Leaves are simple, flowers are unisexual and white in colour, and berry is oblong to ovoid, distributed throughout Amboli Ghat, Maharashtra state, Western Ghats of India.

Ethnopharmacognosy: Our survey revealed that the traditional healers and Dhangars of Ramghat region of western ghat were using this plant to expelling worms, skin diseases, in the treatment of arthritis, dysentery and intestinal infections. From the other species of *Diospyros*, oxygenated-2-naphthaldehydes, naphthoic acids, naphthoates and naphthoquinone dimers have been earlier reported¹⁴⁻¹⁶, with affinities to the African crocodile bark-tree, *D. quiloensis* which has been reported to produce several oxygenated 2-naphthaldehydes¹⁷. To the best of the author's knowledge there has been no report on the phytochemical constituents of this tree.

2. MATERIALS AND METHODS:

General Instrumentation: Melting points were recorded on a Cipla I-28, digital apparatus and were uncorrected. Silica gel (Acme) 60.120 mesh for column chromatography and silica gel (Acme) 100-200 mesh were used for

preparative thin layer chromatography. Spots on chromatogram were detected under UV light and by spraying with 5% H₂SO₄ in methanol. UV spectra were recorded in methanol. Both 1-D and 2-D NMR spectra were run in CDCl₃ (1H: 500 MHz; 13C: 125 MHz) on a Bruker AVANCE DRX-500 spectrometer. Accurate mass measurements were determined on a Kratos M525 RFA instrument. LC/MS (EI) was recorded using an Agilent 1100 series LC/MSD in the APCI mode.

Plant material: The roots of *D. oocarpa* were collected at Amboli ghat, of Maharashtra state, Western ghats of India, The voucher specimen (SGDO-1) was deposited in the College of Pharmaceutical Sciences, Andhra University, Visakhapatnam, India

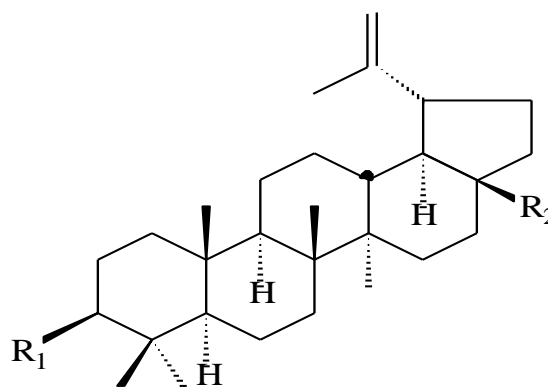
Extraction: Shade dried roots (1.6 kg) were powdered in a Wiley mill and then extracted over four days with chloroform (4 x 1.5 L) at room temperature. TLC examination of the residue showed a number of spots (solvent system: Benzene: Chloroform: Ethyl acetate. 1:3:1). The combined extracts were concentrated under reduced pressure to yield 28 g of reddish brown gum and hence a portion of the chloroform extract (20 g) was chromatographed over silica gel and eluted, in succession, with petroleum ether (b.p 40-60°C), petroleum ether-benzene mixtures containing increasing amounts of benzene, benzene and benzene- Chloroform mixtures containing increasing amounts of chloroform and 250 ml fractions were collected.

Chromatographic separation: TLC examination of the residue showed a number of spots (Benzene: Chloroform: Ethyl acetate. 1:3:1) on spraying with 5% alcoholic H₂SO₄ followed by heating. Column chromatography of the chloroform extract (12 g) over silica gel, on elution with petroleum ether containing increasing amounts of benzene and then Chloroform afforded three coloured bands which yielded pure crystalline material in the sequence DNR-01 to DNR-10

3. RESULTS:

DOR-01 (0.120 g Lupeol)

Crystallized from pet. ether - benzene as colourless needles, mp 212-213°C. [α]_D²⁰ + 240 (CHCl₃, c 0.1) and analysed for the empirical formula C₃₀H₅₀O. It gave a pink colour in L.B reaction and a yellow colour with tetranitromethane. The IR showed absorption bands at 3540(-OH), 1380 and 1390 (gem dimethyl) and at 890 cm⁻¹ (vinyl methylene). The 1H NMR exhibited signals (90 MHz, CDCl₃, δ) at 0.78, 0.80, 0.83, 0.90 and 1.02 (18H, s, 6(CH₃)) 1.63 (3H, s, CH₃-CH=CH₂), 2.25 (1H, d, 19-H), 3.15 (1H, m, 3β-H), 4.51 (2H, d, CH₂



DOR-01 R₁: β-OH., R₂:CH₃

DOR-02 (0.025 g., 5-Hydroxy-4-methoxy-2-naphthaldehyde)

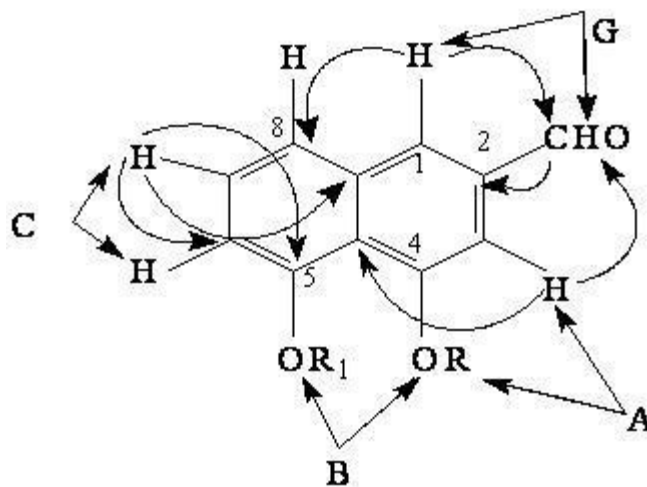
On repeated preparative silica gel TLC using petroleum ether-chloroform (50:50), the upper red band yielded orange-red needles from MeOH, mp 103-104°C.

¹H-NMR (δ CDCl₃, 500.13 MHz) : 7.92 d (1.3) H-1, 7.23 d (1.3) H-3, 7.08 m H-6, 7.48 m H-7/H-8, 10.10 s CHO, 9.30 s 5-OH, 4.14 s 4-OCH₃

¹³CNMR (δ CDCl₃ 125.77 MHz) : 130.7 (C-1), 134.6 (C-2), 98.9 (C-3), 157.5 (C-4), 117.8 (C-4a), 155.0 (C-5), 114.7 (C-6), 129.2 (C-7), 121.0 (C-8), 136.0 (C-8a), 191.9 (CHO), 56.7 (C-4OCH₃)

LC-MS (EIMS) (m/z) (%) : [M+H]⁺ 203 (100%)

Calculated for C₁₂H₁₀O₃ and found C. 71.28%, H. 4.98% and O. 23.74%.



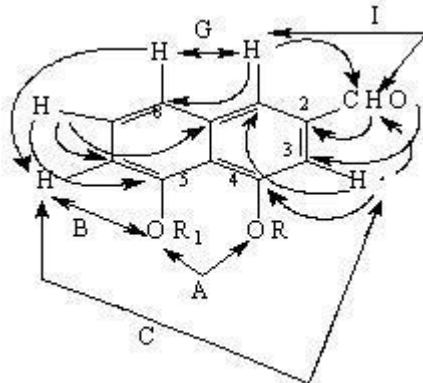
R = CH₃, R₁=H

NOESY and HMBC correlations of DOR-02

DOR-03 (0.030 g, 4-Hydroxy-5-methoxy-2-naphthalde)

On repeated preparative silica gel TLC using petroleum ether-chloroform (50:50), the lower band afforded yellow needles from MeOH, mp 85-90 °C.

UV λ_{max} (log ϵ) : 221 (4.42), 254 (4.49) 379 (3.76)
 $^1\text{H-NMR}$ (δ CDCl_3 , 500.13 MHz) : 7.82 *d* (1.2) H-1, 7.31 *d* (1.2) H-3, 6.96 *d* (7.8) H-6, 7.44 *t* H-7 (7.9), 7.59 *d* (8.3) H-8, 10.07 *s* CHO, 9.44 *s* 4-OH, 4.11 *s* 5-OCH₃
 $^{13}\text{CNMR}$ (δ CDCl_3 , 125.77 MHz) : 125.1 (C-1), 136.0 (C-2), 107.0 (C-3), 155.8 (C-4), 118.4 (C-4a), 156.4 (C-5), 107.4 (C-6), 127.2 (C-7), 123.6 (C-8), 136.3 (C-8a), 192.3 (CHO), 56.6 (C-4OCH₃),
 HR-EIMS (EIMS) (m/z) (%) : 202⁺ (100%)
 C₁₂H₁₀O₃ was found 202.0627, requires 202.0630.



R=H, R₁=CH₃

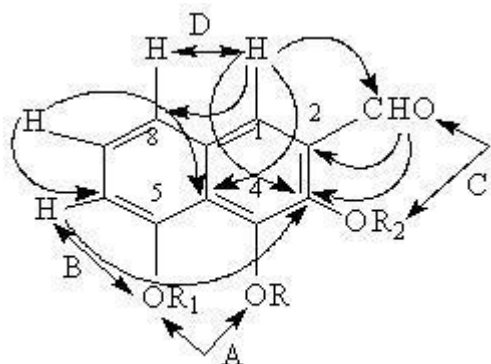
NOESY and HMBC correlation DOR-03

DOR-04 (0.023 g, 4-Hydroxy-3, 5-dimethoxy-2-naphthaldehyde)

On Fractional crystallization using methanol yielded yellow needles, mp 151-15

UV λ_{max} (log ϵ) : 231 (4.95), 260 (4.84), 291 (4.22), 390 (3.88)
 $^1\text{H-NMR}$ (δ CDCl_3 , 500.13 MHz) : 7.85 *s* H-1, 6.89 *d* (7.7) H-6, 7.32 *t* H-7 (7.9), 7.51 *d* (8.3) H-8, 10.53 *s* CHO, 9.46 *s* 4-OH, 4.07 *s* 3-OCH₃, 4.11 *s* 5-OCH₃
 $^{13}\text{CNMR}$ (δ CDCl_3 , 125.77 MHz) : 119.7 (C-1), 129.7 (C-2), 144.1 (C-3), 147.1 (C-4), 119.2 (C-4a), 155.8 (C-5), 106.7 (C-6), 125.7 (C-7), 124.0 (C-8), 132.1 (C-8a), 191.0 (CHO), 62.1 (C-4OCH₃), 56.6 (C-5OCH₃),
 HR-EIMS (EIMS) (m/z) (%) : 232⁺ (100%)

C₁₃H₁₂O₄ was calculated and found 232.0733 and requires 232.0736



R=H., R₁= R₂=CH₃

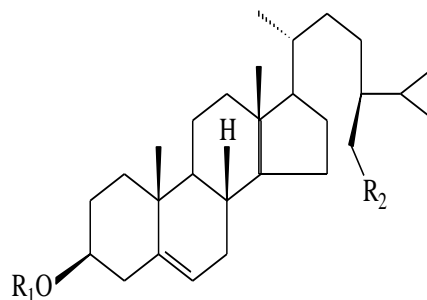
NOESY and HMBC correlations of DOR-04

DOR-05 (0.033g, β -sitosterol)

Crystallized from pet. ether-benzene as colourless needles, mp 134-136 °C, $[\alpha]_D^{30}$ - 36 ° (CHCl_3 , c, 1.01) and had an empirical formulae C₂₉ H₅₀ O and was found to be C, 83.7; H, 12.7 % and requires C, 84.0; H, 12.2 %. The IR

spectrum showed absorption bands IR ν_{max}^{KBR} 3440, 2970, 2910, 2880, 1470, 1385, 1380, 1055 cm^{-1} .

The above data coincided well with that of β -sitosterol and the identity was confirmed by comparison with an authentic sample through m.mp and Co-TLC.



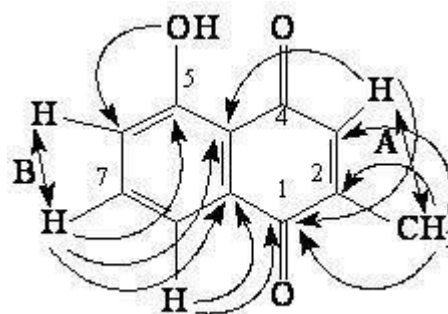
R₁:H., R₂:CH₃

DOR-06 (0.023g, Plumbagin)

On crystallization from methanol, orange needles were obtained. It had an m.p. 78-79° C

IR ν_{max}^{KBR} (cm^{-1}) :1665, 1645 cm^{-1}
 $^1\text{H-NMR}$ (δ CDCl_3 , 500.13 MHz) : 2.20 *d* (1.5) 2-CH₃, 6.81 *d* (1.5) H-3, 11.97 *s* 5-OH, 7.25 *dd* (8.1, 1.4) H-6, 7.60 *br. t* (7.6) H-7, 7.64 *dd* (7.5, 1.4) H-8
 $^{13}\text{CNMR}$ (δ CDCl_3 , 125.77 MHz) :185.0 (C-1), 149.8 (C-2), 16.7 (2-CH₃), 135.7 (C-3), 190.5 (C-4), 161.4 (C-5OH), 124.4 (C6), 136.3 (C-7), 119.5 (C-8), 115.4 (C-4a), 132.3 (C-8a),

Calculated for C₁₁H₈O₃ and found C. 70.2%, H. 4.28% and O. 25.51%.



Selected NOESY and HMBC correlations of DOR-06

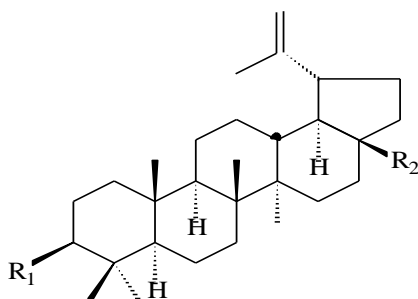
DOR-07 (0.035 g, Betulinaldehyde)

Betulinaldehyde (lupane type) was crystallized from methanol as colourless needles, mp. 185-187° Molecular formula C₃₀H₄₈O₂.

IR ν_{max}^{KBR} 3380, 3060, 2950, 2880, 1730, 1470 cm^{-1} .

Specific rotation $[\alpha]_D^{20} + 27^\circ$ (CHCl₃, c, 1.01) indicating it to be a dextrorotatory compound.

¹H NMR exhibited signals (300 MHz, CHCl₃, δ) at 0.73 (3H, s, 27-Me), 0.80 (3H, s, 25-Me), 0.90 (3H, s, 26-Me), 0.95 (6H, s, 23-Me, 24-Me), 1.68 (3H, s, *br*, 29-Me), 2.80 (1H, *m*, H-19), 3.15 (1H, *dd*, *J* = 11 Hz and 6 Hz, H-3), 4.60, 4.72 (2H, 2 x *s* (*br*) H-30), 9.65 (1H, *s*, H-28). From the above data the compound DAR-07 was identified and confirmed by comparison with an authentic sample through m.m.p. and Co- TLC^{18, 19}



(DOR-07) Betulinaldehyde., R₁: β-OH., R₂: CHO

DOR-08 (0.040g, Diospyrin)

On silica gel eluting with CHCl₃-MeOH (99:1) afforded the less polar orange red prisms, m.p 258 °C. It had an R_f 0.27 in benzene.

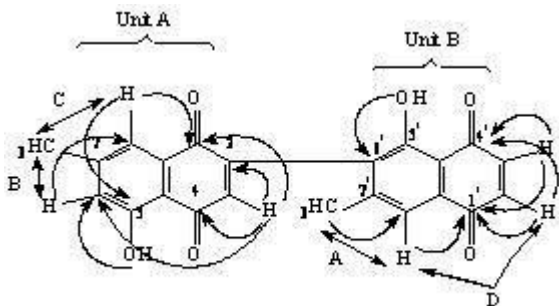
UV λ_{max} nm (log ε) : 252 (4.34), 432 (3.9).

¹HNMR (δ CDCl₃, 500.13 MHz) : 6.91 *s* H-3, 11.90 *s* 5-OH, 7.14 *s* H-6, 2.47 *s* 7-CH₃, 7.51 *s* H-8, 6.96 *s* H-2'/H-3', 12.10 *s* 5'-OH, 2.32 *s* 7'-CH₃, 7.57 *s* H-8'.

¹³CNMR (δ CDCl₃ 125.77 MHz) : 182.8 (C-1), 146.0 (C-2), 139.1 (C-3), 189.1 (C-4), 162.0 (C-OH), 124.5 (C-6), 148.9 (C-7), 22.5 (7-CH₃), 121.5(C-8), 131.9 (C-9), 113.4 (C-10), 184.3 (C-1'), 139.7 (C-2'), 139.0 (C-3'), 190.0 (C-4'), 159.4 (C-OH), 129.1 (C-6'), 146.7 (C-7'), 21.3 (7-CH₃), 121.0 (C-8'), 131.6 (C-9'), 113.2 (C-10').

HR-MS (EIMS) (m/z) (%) : 374 (M⁺ 100%), 359 (20), 328 (14), 187 (16), 163(12), 134 (18), 106 (54) and was found to be a dimer.

It was calculated for C₂₂H₁₄O₆ and found C.70.59%, H. 3.77% and O. 25.64%.



NOESY and HMBC correlations of DOR-08

DOR- 09 (0.070 g, 8'hydroxyisodiospyrin)

On silica gel eluting with CHCl₃-MeOH (99:1) afforded a more polar eluate that on crystallization using benzene yielded 0.070 g of red crystals, m.p 275-277 °C with R_f 0.12 (benzene).

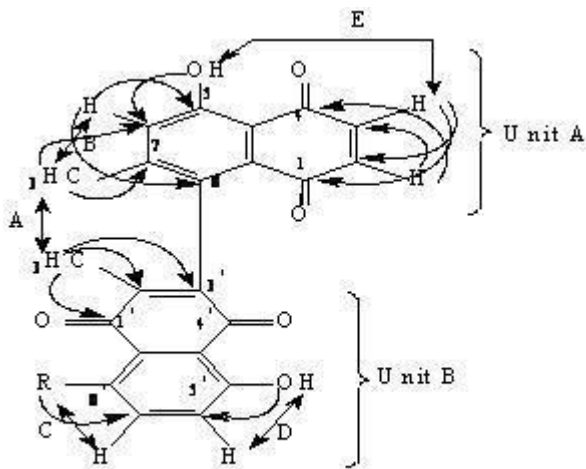
$[\alpha]_D^{23}$ (dioxan, c 0.1) : -72° (laevorotatory)
 UV λ_{max} nm (log ε) : 253 (4.24), 432 (3.84), 460 (3.92), 4.84 (3.91) 554 (3.66)

¹H-NMR (δCDCl₃, 500.13 MHz) : 6.78 *d* (15.7) H-2, 6.95 *d* (15.7), 12.35 *s* 5-OH, 7.30 *m* H-5, 2.20 *s* 7-CH₃, 1.87 *s* 2-CH₃, 12.32 *s* 5'-OH, 7.29 *m* H-6', 7.29 *m* H-7', 12.67 *s* 8'-OH,

¹³CNMR (δ CDCl₃ 125.77 MHz) : 184.9 (C-1), 139.9 (C-2), 138.2 (C-3), 190.1 (C-4), 162.3 (C-5OH), 125.9 (C-6), 146.8 (C-7), 20.8 (7-CH₃), 127.2 (C-8), 129.3 (C-9), 114.2 (C-10), 186.7 (C-1'), 147.3 (C-2'), 13.6 (2'-CH₃), 143.0 (C-3'), 158.9 (C-5'OH), 129.9 (C-6'), 129.9 (C-7'), 158.9 (C-8'OH), 112.3 (C-9').

LC-EIMS, m/z (%) : 390 (M⁺ 100%), 359 (23), 127 (14), 121 (18), 115 (43), 97 (15), 95 (14), 94 (28), 92 (60), 92 (78), 77 (22), 76 (21), 69 (23)

Calculated for C₂₂H₁₄O₇ and found C. 70.59%, H. 3.77% and O. 30.81%.



R=OH

NOESY and HMBC correlation of DOR-09

DOR- 10 (0.124 g, Umbelliferone)

On silica gel eluting with CHCl₃-MeOH (99:1) afforded a more polar eluate that on crystallization using ethylacetate yielded 0.124 g of colourless prisms, m.p 229-231 °C.

Analysed for the Molecular formula C₉ H₆ O₃.

UV λ_{max} : 206, 325 nm.

IR ν cm^{-1} : 3159, 1681, 1620, 1568, 1322, 1235, 1134 and 835 cm^{-1} .
 Mass spectrum at EIMS (70 eV) m/z (%) : 162 (100), 134 (81), 51 (21).
 NMR spectrum (400 MHz, in $\text{DMSO}-d_6$) : δ 6.19 (d, $J=9.4$ Hz, 1H), 7.93 (d, $J=9.4$ Hz, 1H), δ 7.52 (1H, $J=8.4$ Hz, H-5), δ 6.87 (1H, dd, $J=0.2, 8.4$ Hz, H-6) and δ 6.70 (1H, d, $J=0.16$ Hz, H-8)
 ^{13}C NMR spectra (100 MHz, DMSO) : δ 161.3, 144.5, 111.4, 129.7, 113, 1, 160.4, 102.1, 155.5.

4. DISCUSSIONS:

The chemical examination of the roots of *D. oocarpa* afforded ten compounds on column chromatography and repeated crystallizations, Lupeol (**DOR-01**), 5-Hydroxy-4-methoxy-2-naphthaldehyde (**DOR-02**), 4-Hydroxy-5-methoxy-2-naphthaldehyde (**DOR-03**), 4-Hydroxy-3, 5-dimethoxy-2-naphthaldehyde (**DOR-04**), β -sitosterol, (**DOR-05**), Plumbagin (**DOR-06**), Betulinaldehyde (**DOR-07**), Diospyrin (**DOR-08**), 8'-hydroxyisodiospyrin (**DOR-09**), Umbelliferone (**DOR-10**). All these compounds are characterized by conventional chemical tests, physical properties and spectroscopic methods like UV, IR, NMR, ^{13}C NMR and MASS. Out of these compounds, **DOR-1**, **DOR-4** and **DOR-7** belong to triterpene groups and **DOR-2**, **DOR-3**, **DOR-4**, **DOR-6**, **DOR-8** and **DOR-9** are naphthoquinones. The compound **DOR-10** named as umbelliferone, interestingly is a coumarin class. Occurrence of naphthoquinones is very common in *Diospyros* species. The author could isolate these quinones from this species also and thus justified the species. The compound diospyrin is also frequent in the genus *Diospyros* and thus serving as chemotaxonomic marker of the genus. It is also interesting to note that three naphthaldehydes were also isolated from this species. All these three aldehydes, 5-Hydroxy-4-methoxy-2-naphthaldehyde (**DOR-02**), 4-Hydroxy-5-methoxy-2-naphthaldehyde (**DOR-03**), 4-Hydroxy-3, 5-dimethoxy-2-naphthaldehyde (**DOR-04**) are the first time report from this species. Distribution of naphthaldehydes in nature is very rare and this report of occurrence is new to the literature. The compound (**DOR-07**) betulinaldehyde was isolated in very good quantity i.e upto 5% yield.

5. ACKNOWLEDGEMENTS:

We would like to thank ICMR for the financial support for this research (21/12/17/09/HSR, dated: 24/06/2010).

6. REFERENCES:

- 1) Arunendra P, Dines KK, Rakesh M. Phytochemistry 2004; 65:2153.
- 2) Mallavadhani UV, Panda AK, Rao YR. Phytochemistry. 1998; 49(4): 901.
- 3) Ganapaty S, Thomas PS, Fotso S, Laatsch H. Phytochemistry.2004; 65(9): 1265.

- 4) Ganapaty S, Thomas PS, Karagianis G, Peter GW. Nat.Prod.Res. 2006; 20(9): 783.
- 5) Ganapaty S, Thomas PS, Karagianis G, Peter G W, Brun R. Phytochemistry. 2006; 67(17): 1950.
- 6) Rajarajeshwari N, Ganapaty S, Harish Kumar D H. INT.J.PH.SCI.2010; 2(1):445-447.
- 7) Sastry, B.N., 1952. The Wealth of India Raw Materials, Vol. III. CSIR, New Delhi, p. 76.
- 8) Sankaram, A.V.B., Reddy, V.V.N., Phytochemistry. 1984; 23 (9), 2039.2042.
- 9) Christophewiart, Medicinal Plants of Asia and the Pacific, 2006, ed illustrated, CRC publication, 72-81.
- 10) Chopra R.N, Nayar S.L., Chopra I.C. Glossary of Indian Medicinal Plants. 1956, New Delhi, India, CSIR, 98-99.
- 11) Kirtikar K.R., Basu B.D. Indian Medicinal Plants, 1933, In Blatter E, Cains JF, Mhaskar KS (eds). Allahabad, India: Lalit Mohan Basu, 1498.
- 12) Shou-Ming, Z., Waterman P.G., Jeffrey J.A.D. Phytochemistry, 1984, 23, (5), 1067-1072.
- 13) Mbi, C.N., Waterman P.G.J. Pharm. Pharmacol., 1978, 30, 86-88.
- 14) Gupta, R.K. and Mahadevan, V., Indian Journal of Pharmacology, 1967, 29, 289.
- 15) Brown, A.G. and Thomson, R.H., Journal of the Chemical Society, 1965, 4292.
- 16) Sankaram, A.V.B. and Reddy, V.V.N., Phytochemistry, 1984, 23(9), 2039.
- 17) Ganguly A K., and Govindachari T R., Tetrahedron. Letters. 1966, 3373.
- 18) Zhong, S.M., Waterman, P.G. and Jeffrey, J.A.D., Phytochemistry, 1984, 23, 1067
- 19) Chen, C.C., Yu, H.J. and Huang, Y.L., Zhonghua Yaoxue Zazhi, 1992, 44, 229; Chemical Abstracts, 1992, 117, 188243.

Conflict of Interest: None Declared

Cite this article as:

M J Amar dev, N Rajarajeshwari. Phytoconstituents isolated from *Diospyros oocarpa* Thwait. Asian Journal of Biomedical and Pharmaceutical Sciences, 2013, 3: (19), 50-54.